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Kinetically Controlled Photoinduced Electron Transfer Switching in Cu(I)-Responsive Fluorescent Probes

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Abstract: Copper(I)-responsive fluorescent probes based on photoinduced electron transfer (PET) switching consistently display incomplete recovery of emission upon Cu(I) binding compared to the corresponding isolated fluorophores, raising the question of whether Cu(I) might engage in adverse quenching pathways. To address this question, we performed detailed photophysical studies on a series of Cu(I)-responsive fluorescent probes that are based on a 16-membered thiazacrown receptor ([16]aneNS₃) tethered to 1,3,5triarylpyrazoline-fluorophores. The fluorescence enhancement upon Cu(I) binding, which is mainly governed by changes in the photoinduced electron transfer (PET) driving force between the ligand and fluorophore, was systematically optimized by increasing the electron withdrawing character of the 1-aryl-ring, yielding a maximum 29-fold fluorescence enhancement upon saturation with Cu(I) in methanol and a greater than 500-fold enhancement upon protonation with trifluoroacetic acid. Time-resolved fluorescence decay data for the Cu(I)-saturated probe indicated the presence of three distinct emissive species in methanol. Contrary to the notion that Cu(I) might engage in reductive electron transfer quenching, femtosecond time-resolved pump-probe experiments provided no evidence for formation of a transient Cu(II) species upon photoexcitation. Variable temperature ¹H NMR experiments revealed a dynamic equilibrium between the tetradentate NS₃-coordinated Cu(I) complex and a ternary complex involving coordination of a solvent molecule, an observation that was further supported by quantum chemical calculations. The combined photophysical, electrochemical, and solution chemistry experiments demonstrate that electron transfer from Cu(I) does not compete with radiative deactivation of the excited fluorophore, and, hence, that the Cu(I)induced fluorescence switching is kinetically controlled.

Introduction

The development of synthetic fluorescent probes for the detection of transition metal cations represents a very active and steadily growing research field. An important class of cation-responsive probes is based on a photoinduced electron transfer (PET) switching mechanism, which translates the metal-binding event into a fluorescence emission increase. While numerous PET probes have been reported for the detection of closed-shell metal cations, including Zn(II), Cd(II), and Hg(II), the detection of redox-active transition metals such as Cu(I/II) and Fe(II/III) has received less attention. The fluorescence-based detection of these cations is particularly challenging due to interfering metal-mediated quenching pathways, for example through enhanced spin—orbit coupling or competing energy and electron transfer processes. While these

undesired quenching pathways can be reduced or in some cases even eliminated through the use of electronically decoupled metal receptor moieties, ¹⁰ reports on Cu(I)-responsive PET

(5) Prodi, L.; Bargossi, C.; Montalti, M.; Zaccheroni, N.; Su, N.; Bradshaw,

- J. S.; Izatt, R. M.; Savage, P. B. J. Am. Chem. Soc. 2000, 122, 6769. Rurack, K.; Resch-Genger, U.; Bricks, J. L.; Spieles, M. Chem. Commun. 2000, 2103. Nolan, E. M.; Lippard, S. J. J. Am. Chem. Soc. 2003, 125, 14270. Yoon, S.; Albers, A. E.; Wong, A. P.; Chang, C. J. J. Am. Chem. Soc. 2005, 127, 16030. Nolan, E. M.; Racine, M. E.; Lippard, S. J. Inorg. Chem. 2006, 45, 2742. Meng, X. M.; Liu, L.; Hu, H. Y.; Zhu, M. Z.; Wang, M. X.; Shi, J.; Guo, Q. X. Tetrahedron Lett. 2006, 47, 7961. Nolan, E. M.; Lippard, S. J. J. Am. Chem. Soc. 2007, 129, 5910. Yoon, S.; Miller, E. W.; He, Q.; Do, P. H.; Chang, C. J. Angew. Chem., Int. Ed. 2007, 46, 6658. Wang, J.; Qian, X. Org. Lett. 2006, 8, 3721. For a comprehensive review see also: Nolan, E. M.; Lippard, S. J. Chem. Rev. 2008, 108, 3443.
- (6) Rurack, K.; Kollmannsberger, M.; Resch-Genger, U.; Daub, J. J. Am. Chem. Soc. 2000, 122, 968.
- (7) (a) Yang, L.; McRae, R.; Henary, M. M.; Patel, R.; Lai, B.; Vogt, S.; Fahrni, C. J. *Proc. Natl. Acad. Sci. U.S.A.* 2005, 102, 11179. (b) Verma, M.; Chaudry, A. F.; Morgan, M. T.; Fahrni, C. J. *Org. Biomol. Chem.* 2010, 8, 363.
- (8) Cody, J.; Fahrni, C. <u>J. Tetrahedron</u> 2004, 60, 11099. Zeng, L.; Miller, E. W.; Pralle, A.; Isacoff, E. Y.; Chang, C. J. <u>J. Am. Chem. Soc.</u> 2006, 128, 10.
- (9) Fakih, S.; Podinovskaia, M.; Kong, X.; Collins, H. L.; Schaible, U. E.; Hider, R. C. J. Med. Chem. 2008, 51, 4539. Bricks, J. L.; Kovalchuk, A.; Trieflinger, C.; Nofz, M.; Büschel, M.; Tolmachev, A. I.; Daub, J.; Rurack, K. J. Am. Chem. Soc. 2005, 127, 13522.
- (10) Rurack, K.; Resch-Genger, U. Chem. Soc. Rev. 2002, 31, 116.

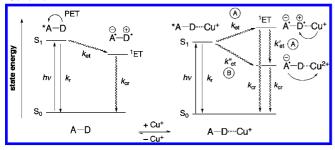
⁽¹⁾ Demchenko, A. P. *Introduction to Fluorescence Sensing*; Springer: New York, 2008.

⁽²⁾ de Silva, A. P.; Gunaratne, H. Q. N.; Gunnlaugsson, T.; Huxley, A. J. M.; McCoy, C. P.; Rademacher, J. T.; Rice, T. E. <u>Chem. Rev.</u> 1997, 97, 1515. Rurack, K. <u>Spectrochim. Acta. Part A</u> 2001, 57, 2161.

Nolan, E. M.; Lippard, S. J. Acc. Chem. Res. 2009, 42, 193. Jiang,
 P. J.; Guo, Z. J. Coord. Chem. Rev. 2004, 248, 205. Kikuchi, K.;
 Komatsu, K.; Nagano, T. Curr. Opin. Chem. Biol. 2004, 8, 182.

⁽⁴⁾ Cheng, T. Y.; Xu, Y. F.; Zhang, S. Y.; Zhu, W. P.; Qian, X. H.; Duan, L. P. J. Am. Chem. Soc. 2008, 130, 16160. Xue, L.; Liu, C.; Jiang, H. Org. Lett. 2009, 11, 1655.

Scheme 1



probes have consistently shown incomplete recoveries of the fluorescence quantum yields compared to the corresponding isolated fluorophores, thus raising the question of whether Cu(I) might be directly involved in adverse quenching pathways. Here we report on detailed photophysical studies that were aimed at clarifying the role of Cu(I) in mediating fluorescence quenching in PET probes.

The molecular architecture of PET probes is based on a donor-spacer-acceptor (D-A) framework that combines a cation receptor with a suitable fluorophore. In the absence of the analyte, the fluorescence emission is strongly quenched due to a fast intramolecular photoinduced electron transfer reaction from the electron-rich cation receptor (D) to the excited fluorophore (A*) acting as the electron acceptor (Scheme 1, left). The formed radical ion pair (D⁺-A⁻) undergoes nonradiative charge recombination back to the initial ground state. Binding of the metal cation to the receptor moiety results in a less favorable donor potential, which slows down the kinetics for PET quenching and thus increases the emission quantum yield. Because redox-active cations such as Cu(I) may also act as electron donors, two additional nonradiative deactivation pathways A and B must be considered (Scheme 1, right). In pathway A, an analogous radical ion pair (D^+-A^-) is formed first, albeit with a slower rate due to an increase in donor potential in the presence of Cu(I), followed by an electron transfer from Cu(I) to the radical cation (D*+). Back-electron transfer to the intermediately formed Cu(II) species then regenerates the initial ground state. Because electron donation from Cu(I) is preceded by formation of the radical ion pair, only the rate constant $k_{\rm et}$ of the first ET reaction will affect the emission response. Hence, Cu(I) would appear to act as a Lewis acid, and in principle, it should be possible to fully recover the fluoresence emission of the unquenched fluorophore. In contrast, pathway B directly competes with radiative deactivation to form the same Cu(II)intermediate (Scheme 1, right). Depending on the rate constant k''_{et} , this pathway might severely compromise the fluorescence recovery, even if $k_{\rm et}$ is much slower than $k_{\rm r}$ in the presence of Cu(I).

While any Cu(I)-responsive PET probe with D-A architecture would be suitable to explore the importance of these pathways, we decided to take advantage of the synthetic accessibility and photophysical tunability of 1,3,5-triarylpyrazoline-based probes (Chart 1a). In these fluorophores, two of the key PET parameters, the excited-state equilibrium energy ΔE_{00} and the acceptor potential $E(A/A^-)$, can be rationally tuned to adjust the change in free energy of the PET reaction ($\Delta G_{\rm et}$)

for a given donor potential $E(D^+/D)$ as expressed by the Rehm-Weller equation, eq 1.¹⁴

$$\Delta G_{\text{et}} = E(D^{+}/D) - E(A/A^{-}) - \Delta E_{00} + w_{\text{p}}$$
 (1)

The term w_p refers to the Coulombic stabilization energy of the radical ion pair formed upon PET. In polar solvents, this value is typically small due to dielectric screening.

The fluorophore π -system in 1,3,5-triarylpyrazolines is composed of the central pyrazoline core and the two aryl rings in the 1- and 3-positions, which are coupled through the sp²hybridized nitrogen and carbon atoms of the heterocycle. Electron withdrawing substituents R² on the aryl ring in the 1-position of the pyrazoline system yield a large increase of ΔE_{00} but only a small change of $E(A/A^{-})$. Conversely, substituents R^1 on the 3-aryl ring influence $E(A/A^-)$ more strongly than ΔE_{00} . This property can be exploited to gradually adjust the PET driving force for any given receptor unit and thus to optimize fluorescence enhancement upon analyte binding. 12 The aryl ring in the 5-position is electronically decoupled from the fluorophore π -system, thus offering an opportunity to attach a metal-receptor R³ for PET switching. ⁷ To bind Cu(I) with high affinity and sufficient redox stability, ¹⁵ we chose a 16-membered trithiazacyclohexadecane ([16]aneNS₃) macrocycle as the receptor moiety (Chart 1b). Furthermore, previous studies showed that the donor potential of N,N-dialkylanilines is well matched with the acceptor potential of pyrazolines carrying a p-cyano substituent on the 3-aryl ring. 12 On the basis of these considerations, we synthesized a series of pyrazoline derivatives 1a-e with varying PET driving forces and evaluated their photophysical properties. To gauge the degree of fluorescence recovery upon saturation with Cu(I), we also characterized each derivative in acidic solution. Because the protonated anilinium cation is electrochemically inactive, acidic conditions should lead to a full recovery of the fluorescence emission. 12 Finally, we performed detailed time-resolved fluorescence and transient absorption experiments to elucidate the role of the proposed excited-state pathways.

Experimental Section

Synthesis. Procedures for the synthesis of derivatives **1a**-**e** and all intermediates are provided in the Supporting Information.

Steady-State Absorption and Fluorescence Spectroscopy. All sample solutions were filtered through 0.2 μ m Teflon membrane filters to remove interfering dust particles or fibers. UV-vis absorption spectra were recorded at 25 °C using a Varian Cary Bio50 UV—vis spectrometer with constant-temperature accessory. Steady-state emission and excitation spectra were recorded with a PTI fluorimeter. For all measurements the path length was 1 cm with a cell volume of 3.0 mL. The fluorescence spectra have been corrected for the spectral response of the detection system (emission correction file provided by instrument manufacturer) and for the spectral irradiance of the excitation channel (via a calibrated photodiode). Fluorescence titrations with Cu(I) were carried out at a fluorophore concentration of 2.3 μ M in MeOH by adding 0.1 molar equiv from a 1 mM stock solution of [Cu(I)(CH₃CN)₄]PF₆ in MeOH containing 10% CH₃CN (v/v). Because the p K_a of trifluoroacetic acid (TFA) is approximately 5 p K_a units higher in methanol compared to aqueous solution, ¹⁶ a large excess of TFA (180 mM) was required to ensure quantitative protonation of the aniline nitrogen as determined by saturation experiments. Quantum yields

⁽¹¹⁾ Verma, M.; Chaudhry, A. F.; Fahrni, C. <u>J. Org. Biomol. Chem.</u> 2009, 7, 1536.

⁽¹²⁾ Cody, J.; Mandal, S.; Yang, L.; Fahrni, C. J. J. Am. Chem. Soc. 2008, 130, 13023.

⁽¹³⁾ Fahrni, C. J.; Yang, L. C.; VanDerveer, D. G. <u>J. Am. Chem. Soc.</u> 2003, 125, 3799.

⁽¹⁴⁾ Rehm, D.; Weller, A. Isr. J. Chem. 1970, 8, 259.

⁽¹⁵⁾ Addison, A. W. *Inorg. Chim. Acta* **1989**, *162*, 217.

⁽¹⁶⁾ Rived, F.; Rosés, M.; Bosch, E. Anal. Chim. Acta 1998, 374, 309.

Chart 1

were determined using quinine sulfate dihydrate in 1.0 N H₂SO₄ as the fluorescence standard ($\Phi_{\rm f} = 0.54 \pm 0.05$). ¹⁷

Picosecond Fluorescence Spectroscopy. Time-resolved fluorescence decay data were acquired at an emission wavelength of 480 nm with a sample concentration of 0.8 μM using a single photon counting spectrometer (Edinburgh Instruments, LifeSpec Series) equipped with a pulsed laser diode as the excitation source (372 nm, fwhm = 80 ps, 10 MHz repetition rate, 1024 channel resolution). Sample solutions of the fluorophore saturated with excess Cu(I) were prepared on the basis of steady-state fluorescence titrations as described above. For measurements under acidic conditions, trifluoroacetic acid was directly added to the sample solution in MeOH (final concentration 180 mM). The time decay data were analyzed by nonlinear least-squares fitting with deconvolution of the instrumental response function using the FluoFit software package.18

Femtosecond Transient Absorption Spectroscopy. Spectra were acquired using a commercial transient absorption spectroscopy system (Newport, Helios). This system accepts two input laser beams, one of variable wavelength used as the pump beam and one of a fixed wavelength (800 nm) used to generate the probe beam in a proprietary nonlinear optical crystal. For the experimental pump beam, the light source was an ultrafast optical parametric amplifier (Newport, TOPAS) running at a repetition rate of 1 kHz, pumped by a Ti:Sapphire regenerative amplifier (Newport, Spitfire). All experiments were performed using the TOPAS output at 350 nm with a pulse width of approximately 120 fs. Approximately 5% of the Spitfire fundamental at 800 nm was used for white light generation (420-800 nm) in the Helios crystal to provide the experimental probe beam. At these specifications, the instrument response function is approximately 200 fs. The time window that can be studied is 3200 ps; the transient spectra were acquired starting at a delay of -1 ps and continued to a delay of 3000 ps. At each time point, data were averaged for 1.5 s. The Helios pump beam was chopped at 500 Hz to obtain pumped (signal) and nonpumped (reference) absorption spectra of the sample. A correction factor for chirp in the probe beam was generated using the ultrafast response of CS2, and was applied to all data sets. The data were stored as 3-D wavelength-time-absorbance matrices that were exported for use with the fitting software. The solutions studied had an OD₃₅₀ in 2 mm path-length cuvettes of 0.07-0.45, and were stirred continually throughout the data acquisition.

Cyclic Voltammetry. The cyclic voltammograms were acquired in methanol or acetonitrile containing 0.1 M Bu₄NPF₆ as the electrolyte using a CH-Instruments potentiostat (model 600A). The samples were measured under inert gas at concentrations ranging from 50 μ M to 3 mM in a single compartment cell with a glassy carbon working electrode and a Pt counter electrode. An aqueous Ag/AgCl reference electrode (3 M KCl) was used in methanol, and a nonaqueous Ag/AgNO₃ (10 mM in 0.1 M Bu₄NPF₆/CH₃CN) reference electrode in acetonitrile. The half-wave potentials were referenced to ferrocene as the internal or external standard as

Computational Methods. All calculations were carried out with the Q-Chem electronic structure calculation suite of programs.²⁰ Ground-state geometries were energy minimized by DFT with the B3LYP hybrid functional²¹ and Pople's 6-31G(d) split valence basis set with added polarization functions. The starting geometry for the Cu(I) complex of ligand 3 (N-phenyl-1,5,9-trithia-13-azacyclohexadecane) was constructed from the crystal structure coordinates $of \ [1,5\text{-}dithia-9,13\text{-}diazacyclohexadecane-copper}(I)] tetrafluoroborate$ (Cambridge crystallographic database, code VEMWAL).²² To ensure a stationary point on the ground-state potential surface, all geometry optimized structures were verified by a vibrational frequency analysis. For the calculation of gas phase reaction enthalpies, single point energies were calculated at the B3LYP/6-31+G(d,p) level of theory and corrected with the respective scaled harmonic zero-point vibrational energies (ZPVE) obtained from the frequency analysis at the B3LYP/6-31G(d) level (scaling factor 0.9806).²³ Molecular orbitals were visualized with the software VMD²⁴ using the Q-Chem plot output data. Details of the computational results including the coordinates of the geometryoptimized structures are provided in the Supporting Information.

Results and Discussion

Contrast Optimization. Following the previously delineated strategy, 11,12 we optimized the contrast ratio upon Cu(I)-binding by progressively increasing the electron withdrawing ability of the 1-aryl ring through fluoro-substitution. All pertinent photophysical data including estimations of the PET driving force for each derivative are provided in Table 1. With increasing number of fluoro substituents, the absorption and emission spectra shift to higher energies while maintaining a uniform band shape across all derivatives (Figure 1). The resulting increase of the corresponding zero-zero transition energies ΔE_{00} , estimated as the average between the absorption and emission maxima, directly parallels the increase in the PET driving force $(-\Delta G_{00})$, which in turn is responsible for the progressively decreasing quantum yields in neutral MeOH (Table 1). For derivative 1e the emission intensities and the associated signal/noise ratios were too low for accurate quantum yield determinations. Upon addition of excess [Cu(I)(CH₃CN)₄]PF₆, all derivatives exhibited substantial fluorescence emission increases. Following the trend of the previous model study, 12 the emission enhancement factors f_e , defined as

recommended by IUPAC. 19 Measurements were typically performed with a scan rate of 100 mV s⁻¹ or as indicated in the text.

⁽¹⁷⁾ Demas, J. N.; Crosby, G. A. J. Phys. Chem. 1971, 75, 991.

⁽¹⁸⁾ Enderlein, J.; Erdmann, R. Opt. Commun. 1997, 134, 371.

⁽¹⁹⁾ Gritzner, G.; Kuta, J. Pure Appl. Chem. 1982, 54, 1527.

⁽²⁰⁾ Shao, Y. E. A.; et al. *Phys. Chem. Chem. Phys.* 2006, 8, 3172.
(21) Becke, A. D. *J. Chem. Phys.* 1993, 98, 5648. (a) Lee, C. T.; Yang,

W. T.; Parr, R. G. Phys. Rev. B 1988, 37, 785. (22) Balakrishnan, K. P.; Riesen, A.; Zuberbühler, A. D.; Kaden, T. A. Acta Crystallogr., Sect. C: Cryst. Soc. Commun. 1990, 46, 1236. Scott, A. P.; Radom, L. J. Phys. Chem. 1996, 100, 16502.

⁽²⁴⁾ Humphrey, W.; Dalke, A.; Schulten, K. J. Mol. Graphics 1996, 14,

Table 1. Photophysical Data of Pyrazoline Derivatives 1a-e in Methanol at 298 K

compd	abs λ_{max} (nm)	em λ_{max} (nm)	Stokes shift (cm ⁻¹)	$\Delta E_{00} \; ({ m eV})^a$	$\Delta G_{ m et}~({ m eV})^b$	$\Phi_{F}{}^c$			$f_{\rm e}{}^d$	
						neutral ^e	acidic f	Cu(I) ^g	acidic f	Cu(I) ^g
1a	394	487	4850	2.85	-0.22	0.0072	0.53	0.15	74	21
1b	381	480	5410	2.92	-0.31	0.0033	0.54	0.095	164	29
1c	373	464	5260	3.00	-0.39	0.0024	0.60	0.048	250	20
1d	356	445	5620	3.13	-0.47	0.0010	0.55	0.020	550	20
1e	350	448	6250	3.15	-0.54	n.d. ^h	0.46	0.020	n.d. ^h	$n.d.^h$

^a Zero-zero transition energy, estimated on the basis of $\Delta E_{00} = (E_{\rm abs}({\rm max}) + E_{\rm cm}({\rm max}))/2$. ^b PET free energy change calculated according to eq 1 with $w_{\rm p} = -0.045$ eV;¹² the experimental donor and acceptor potentials are provided in the Supporting Information. ^c Fluorescence quantum yield, quinine sulfate as reference. ^d Fluorescence enhancement factor $f_{\rm e} = \Phi_{\rm F}/\Phi_{\rm neutral}$. ^e Probe in methanol alone. ^f 180 mM TFA in methanol. ^g 10 μM [Cu(I)(CH₃CN)₄]PF₆ in methanol (0.1% acetonitrile). ^h Signal/noise ratio insufficient for accurate determination.

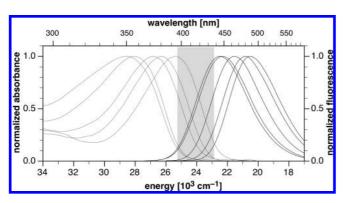


Figure 1. Normalized absorption and emission spectra of pyrazoline derivatives $\mathbf{1a-e}$ in methanol (298 K). Absorption traces are depicted with dotted lines (left) and emission spectra with solid lines (right). The shaded area indicates the tunable range of the excited-state energy ΔE_{00} within each compound series.

the ratio of the quantum yields in the presence and absence of Cu(I), first increased before decreasing with increasing PET driving force, reaching a maximum of 29-fold enhancement for the difluoro-substituted derivative **1b**. In contrast, protonation of the aniline nitrogen under acidic conditions resulted in substantially greater enhancement factors across all derivatives (Table 1), raising the question of whether Cu(I) might be involved in mediating fluorescence quenching. To address this issue, we systematically investigated both the solution chemistry as well as the excited-state dynamics of this system.

Solution Equilibrium with [Cu(I)(CH₃CN)₄]PF₆. The presence of multiple coordination equilibria could complicate the fluorescence response of the probes upon addition of Cu(I). Of particular concern was the possibility of Cu(I)-mediated formation of dimers or higher oligomers, which might undergo selfquenching and thus lower the quantum yield and contrast ratio compared with the acidic solution. To investigate the ligand—Cu(I) binding stoichiometry in methanol, we first performed a molarratio titration with fluorophore 1a and [Cu(I)(CH₃CN)]₄PF₆. As illustrated in Figure 2, the fluorescence emission of 1a increased linearly and was saturated at 1 molar equiv. Excess Cu(I) did not lead to further changes of the emission intensity. In agreement with this observation, a ¹H NMR titration of the isolated ligand N-phenyl-1,5,9-trithia-13-azacyclohexadecane (3) with [Cu(I)(CH₃CN)₄]PF₆ also indicated saturation at 1 molar equiv of Cu(I) (Figure S1, Supporting Information). At substoichiometric concentrations of Cu(I), only single resonances for each set of equivalent protons were observed, indicating rapid metal exchange kinetics exceeding the fast exchange limit on the NMR time scale. The solution chemistry studies thus confirmed the presence of a defined 1:1 ligand—Cu(I) equilibrium and provided no evidence for formation of aggregates or oligomeric coordina-

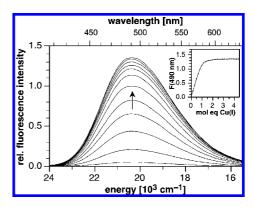


Figure 2. Fluorescence titration of pyrazoline **1a** $(2.3 \mu M)$ with $[Cu(I)(CH_3CN)_4]PF_6$ in MeOH (298 K, excitation at 345 nm). Inset shows a molar-ratio plot for the fluorescence intensity change at 490 nm.

tion species that might compromise the quantum yield and lower the fluorescence enhancement factors compared to the acidic solution.

Copper Redox Chemistry. To study the electrochemical properties of the Cu(II/I) couple coordinated to the thiazacrown receptor without interference from the pyrazoline fluorophore, we utilized the truncated ligand 3 as a model compound. The formal potential of the Cu(II/I) couple coordinated to the ligand can be readily measured by cyclic voltammetry of the Cu(II) complex.²⁵ To determine the amount of Cu(II) needed to saturate ligand 3, we first performed a molar-ratio titration with Cu(OTf)₂ in methanol. Only at a large excess of Cu(II) were we able to discern changes in the UV-vis absorption spectrum, indicating a very low affinity of 3 toward Cu(II). In the presence of 10 mM Cu(OTf)₂, we observed formation of a new band with a maximum at 478 nm (Figure S3a, Supporting Information). Unlike the broad shape expected for a charge-transfer band, the absorption trace showed distinct vibronic structure that resembled the spectrum of the N,N,N',N'-tetramethylbenzidine radical cation, 26 apparently indicating oxidative dimerization of the ligand. Treatment of the isolated dimer (6, see Supporting Information) with Cu(OTf)₂ yielded the same spectral signature at 478 nm, thus independently confirming the formation of the dimer radical cation (Figure S3b, Supporting Information). In contrast, no changes in the UV-vis spectrum were observed for compound 1a under the same conditions (data not shown), indicating that the pyrazoline substituent in the para position

⁽²⁵⁾ Kakos, S. H.; Dressel, L. T.; Bushendorf, J. D.; Kotarba, C. P.; Wijetunge, P.; Kulatilleke, C. P.; McGillivary, M. P.; Chaka, G.; Heeg, M. J.; Ochrymowycz, L. A.; Rorabacher, D. B. <u>Inorg. Chem.</u> 2006, 45, 923.

⁽²⁶⁾ Shida, T. Electronic Absorption Spectra of Radical Ions; Elsevier: Amsterdam, 1988.

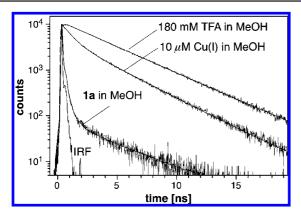


Figure 3. Time-dependent fluorescence decay profile of **1a** in methanol, and in the presence of $10 \,\mu\text{M}$ [Cu(I)(CH₃CN)₄]PF₆, or $180 \,\text{mM}$ trifluoroacetic acid (TFA). A $1 \,\mu\text{M}$ solution of **1a** was excited at 372 nm (80 ps fwhm), and the emission signal was detected by single photon counting at 480 nm. Nonlinear least-squares fitted profiles are shown as solid lines (see Table 2 for fitting data; IRF = instrument response function).

of the aniline ligand inhibits dimerization. While Cu(II)mediated oxidative coupling of N,N-dialkylanilines has been reported in the literature, 27,28 the reaction required acetonitrile, a solvent that greatly favors the Cu(I) state. ²⁹ A formal potential of 0.40 V versus SHE has been reported for the Cu(II/I) couple solvated in methanol.²⁹ To compare this value with the donor potential of ligand 3 measured against Fc+/0 as the reference, the following conversion can be applied. According to the manufacturer, the potential of the aqueous Ag/AgCl (3 M KCl) reference electrode used in our measurements is 0.21 V versus SHE. Given the half-wave potential of 0.38 V of the Fc^{+/0} couple under the experimental conditions, we calculated a corrected potential of -0.19 V versus Fc^{+/0} for the solvated Cu(II/I) couple in methanol. While there remains some uncertainty in the above conversion due to the exclusion of junction potentials, the solvated Cu(II/I) couple is clearly a much weaker oxidant by approximately 0.58 V compared to the radical cation of ligand 3 $(E(D^+/D) = 0.39 \text{ vs Fc}^{+/0})$. Because formation of the latter species is a prerequisite for the observed oxidative coupling reaction, we can conclude that the Cu(II) complex of 3 must act as an oxidant. This in turn implies that the reduction potential of this complex is similar or greater than the estimated donor potential of 0.39 V (vs Fc^{+/0}) of the free ligand 3. Concluding from the electrochemical properties of the Cu(II/I) couple coordinated to the thiazacrown receptor, a competing PET quenching pathway mediated by Cu(I) is thermodynamically

Fluorescence Decay Kinetics. To gain information about the excited-state dynamics and possibly the role of Cu(I) in fluorescence deactivation, we performed time-resolved fluorescence decay measurements with derivative **1a** as a representative compound. The compound was excited at 372 nm with a pulsed laser (80 ps fwhm), and the emission decay signal was collected by single photon counting at 480 nm. As illustrated in Figure 3, in neutral methanol the fluorescence emission decayed rapidly involving complex kinetics. The data fit best with a triexponential decay involving two short components with 34 and 232 ps lifetimes, and a component with 3.53 ns lifetime (Table 2).

Repeated HPLC purification of 1a resulted in a decrease of the long-lifetime component, indicating the presence of a difficult to remove impurity (<0.1%). Despite the small fractional contribution of the long-lifetime component, the triexponential fit yielded greatly improved residuals compared to a biexponential decay model ($\chi^2 = 2.56$).

Upon addition of trifluoroacetic acid (TFA), monoexponential decay kinetics with a lifetime of 3.76 ns were observed. For comparison, we also measured the decay kinetics of compound 2, which contains the same pyrazoline fluorophore but lacks the thiazacrown moiety. In neutral methanol, 2 displayed monoexponential decay kinetics with a lifetime of 3.76 ns. Consistent with effective inhibition of the PET quenching process, under acidic conditions compound 1a yielded indistinguishable decay kinetics compared with 2 in neutral methanol.

Saturation of 1a with Cu(I) in methanol resulted in surprisingly complex decay kinetics. Nonlinear least-squares fitting revealed three distinct decay components with similar fractional amplitudes but distinctly different lifetimes of 84 ps, 0.77 ns, and 3.23 ns (Table 2), suggesting the presence of three distinct emissive species emitting with different decay kinetics. It is noteworthy that the long-lifetime component decays with kinetics comparable to the acidic conditions. The species responsible for this component must therefore invoke similar reduction in the PET quenching kinetics compared to the protonated form. In agreement with weak Cu(II)-binding, addition of $10~\mu M$ Cu(II)(OTf)₂ to a solution of 1a in methanol had no effect on the fluorescence decay kinetics.

Steady-State Fluorescence Emission Profiles. To investigate whether divergent nonequilibrating excited-state pathways might be responsible for the complex decay kinetics, we acquired the fluorescence emission spectrum of 1a saturated with Cu(I) and also under acidic conditions over a broad excitation energy range of 22 000–33 000 cm⁻¹ (Figure 4). Contour plots of the emission spectra revealed symmetrical profiles with a single emission maximum centered at 487 nm (20 530 cm⁻¹) for both conditions. Given the nearly indistinguishable 2D profiles of the protonated versus Cu(I)-saturated probe, we can conclude that the nature of the emissive state is very similar for both conditions. Excited-state processes involving multiple non-equilibrated deactivation pathways can therefore be ruled out in explaining the observed triexponential decay kinetics in the presence of Cu(I).

Transient Absorption Spectroscopy. While picosecond fluorescence decay measurements can provide insights into the excited-state dynamics, the method is not suitable to identify the transient reaction products. To gain further information regarding the excited-state intermediates formed in the presence of Cu(I) compared to neutral or acidic conditions, we performed a series of ultrafast pump-probe experiments with derivative 1a. Of particular interest was the detection of the charge separated radical ion pair formed through the PET reaction. Transient absorption spectra were recorded at different delay times after photoexcitation at 350 nm with a femtosecond laser pulse (120 fs fwhm). To visualize the evolution of the difference spectra over the entire time course, we constructed a set of colorcoded contour plots (Figure 5). Nonlinear least-squares fitting of the time-dependent absorption traces over the entire spectral range yielded deconvoluted spectra for each transient species (Figure 6). All pertinent numerical data including fitting results are compiled in Table 3.

In methanol alone, two distinct regions at 480 and 630 nm (dashed circles) appeared in the difference spectra after a delay

⁽²⁷⁾ Kirchgessner, M.; Sreenath, K.; Gopidas, K. R. J. Org. Chem. 2006, 71, 9849.

⁽²⁸⁾ Sumalekshmy, S.; Gopidas, K. R. *Chem. Phys. Lett.* 2005, 413, 294.
(29) Milazzo, G.; Caroli, S. *Tables of Standard Electrode Potentials*; Wiley: New York, 1978.

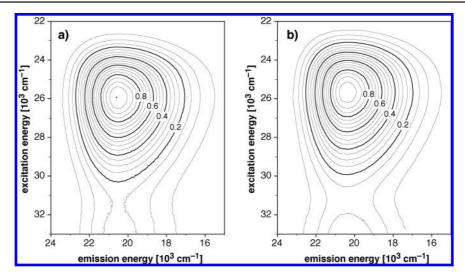


Figure 4. Two-dimensional fluorescence excitation—emission contour plots for 1a in methanol containing $10 \ \mu M \ [Cu(I)(CH_3CN)_4]PF_6$ (a) or $180 \ mM$ trifluoroacetic acid (b).

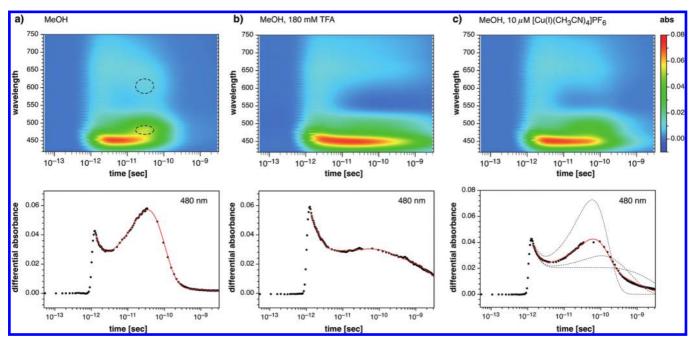


Figure 5. Transient absorption spectra of pyrazoline 1a in (a) methanol, (b) 180 mM TFA in MeOH, and (c) 10μ M [Cu(I)(CH₃CN)₄]PF₆ in MeOH. Top row shows color-coded contour plots of the transient difference absorption traces on a logarithmic time scale. Each plot is represented on the absorption scale shown to the right. Bottom row shows transient absorption progression at 480 nm.

Table 2. Time-Resolved Fluorescence Decay Data for Compounds 1a and 2 in Methanol at 298 K

cmpd	conditions	$ au_i$ (ns) a	a_i^b	f _i c	χ^{2d}	$k_0 (s^{-1})^e$
1a	МеОН	0.0341 ± 0.0001	0.967	0.740	0.851	2.9×10^{10}
		0.232 ± 0.011	0.032	0.167		4.3×10^{9}
		3.53 ± 0.05	0.001	0.093		2.8×10^{8}
1a	TFA (180 mM) in MeOH	3.76 ± 0.01	1.000	1.000	1.145	2.7×10^{8}
1a	$[Cu(I)(CH_3CN)_4]PF_6$ (10 μ M) in MeOH	0.084 ± 0.03	0.347	0.023	0.970	1.2×10^{10}
		0.774 ± 0.007	0.359	0.221		1.3×10^{9}
		3.23 ± 0.04	0.294	0.756		3.1×10^{8}
2	МеОН	3.76 ± 0.01	1.000	1.000	1.380	2.7×10^{8}

^a Fluorescence lifetime. ^b Fractional amplitudes $a_i = \alpha_i / \sum_{i=1}^n \alpha_i$ with $\sum_{i=1}^n \alpha_i = 1$ for the multiexponential decay function $I(t) = \sum_{i=1}^n \alpha_i \exp{(-t/\tau_i)}$. ^c Fractional intensities $f_i = \alpha_i \tau_i / \sum_{i=1}^n \alpha_i \tau_i$. Goodness of fit parameter. ^e Excited-state deactivation rate $k_0 = 1/\tau_i$.

time of approximately 30 ps. The corresponding changes were less pronounced in the presence of Cu(I) and entirely absent under acidic conditions. Given the absorption maximum of 470 nm of the structurally related *N*,*N*-dimethylaniline radical

cation, ^{28,30} the 480 nm band can be assigned to the thiazacrown substituted 5-aryl radical cation formed during the course of

⁽³⁰⁾ Maroz, A.; Hermann, R.; Naumov, S.; Brede, O. <u>J. Phys. Chem. A</u> 2005, 109, 4690.

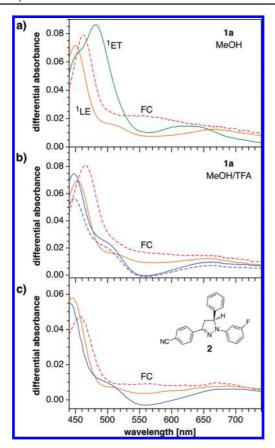


Figure 6. Deconvoluted difference absorption spectra for the transient excited-state species of 1a in (a) methanol and (b) methanol containing 180 mM TFA. (c) Difference absorption spectra for the transient excitedstate species of 2 lacking the PET donor. Legend: FC = Franck-Condon state (dashed red), ¹LE = vibrationally cooled, local excited state (solid yellow), ¹ET = nonemissive singlet electron transfer state (solid green), ¹CT = emissive singlet charge transfer state (blue).

the PET process. The transient absorption progression at 480 nm highlights the differences in evolution of the radical cation in neutral methanol compared to the other two conditions (Figure 5, bottom). The difference spectra fit well with a model composed of a series of four distinct species linked by three consecutive first-order rate constants:

$$FC \xrightarrow{k_1} {}^{1}LE \xrightarrow{k_2} {}^{1}ET \xrightarrow{k_3} GS$$
 (2)

The initially formed Franck-Condon (FC) state decayed with a time constant of 0.6 ps to a local excited state (¹LE), which was converted to the electron-transfer state (¹ET) with a time constant of 24 ps. The characteristic 480 and 630 nm bands of the ¹ET state rapidly decayed with a 77 ps time constant back to the initial ground state (GS).

Consistent with efficient inhibition of PET, the transient absorption traces of 1a in acidic methanol did not exhibit bands at 480 and 630 nm characteristic of the ¹ET state (Figure 5b). The absorption traces were fit again with a model that included five species and four consecutive first-order rate constants:

$$FC \xrightarrow{k_1} {}^{1}LE \xrightarrow{k_2} {}^{1}CT * \xrightarrow{k_3} {}^{1}CT \xrightarrow{k_4} GS$$
 (3)

The deconvoluted difference spectra of the first two intermediates strongly resemble the spectra of the Franck-Condon and local excited states under neutral conditions, which indicates formation of qualitatively similar species (Figure 6a,b). Vibrational cooling from FC to ¹LE again occurred with a fast time constant of 1.1 ps. Interestingly, ¹LE did not immediately decay to the ground state So, but was rapidly converted to two additional intermediate species with qualitatively similar difference absorption spectra (Figure 6b). Using a simplified model composed of a single intermediate species, an average time constant of 90 ps was obtained; however, the fit significantly improved with the expanded model to give two closely spaced time constants of 71 and 110 ps. The latter agreed well with the slightly delayed fluorescence emission already observed in the picosecond lifetime measurements. Given the similarity between the deconvoluted difference spectra, the two intermediate species must have a closely related electronic structure. To explore the potential role of TFA in the delayed fluorescence emission, we also conducted pump-probe experiments in neutral methanol with pyrazoline 2, which lacks the thiazacrown receptor. The deconvoluted transient absorption traces agreed well qualitatively with the deconvoluted spectra of 1a under acidic conditions (Figure 6c); however, the data better matched the simpler four species model lacking the second intermediate. While the pump-probe experiments indicate that strong acidic conditions impose some effect on the excited-state dynamics, there is no substantive evidence for the formation of a structurally different intermediate. Earlier reports on the excitedstate dynamics of pyrazolines concluded that the emissive state is not identical with the initial photoexcited singlet state.³¹ Gas phase quantum chemical calculations on a broad range of pyrazoline fluorophores uniformly identified a strongly allowed lowest excited state S_1 with $\pi - \pi^*$ character, 13 making the involvement of higher lying excited states along with slow internal conversion processes quite unlikely. Consistent with the broad unstructured emission of 1a reminiscent of a polarized charge-transfer state ¹CT, vertical excitation into S₁ is presumably followed by substantial structural reorganization combined with solvent relaxation, both of which may contribute to the delayed radiative deactivation.³² The polarized nature of the emissive states is also in agreement with previous solvatochromic studies employing the Lippert-Mataga formalism, which revealed a substantially increased excited-state dipole moment compared to the ground state with $\Delta\mu_{\rm ge}$ ranging between 8 and 13 D for a broad range of substituted 1,3,5-triarylpyrazolines. 13,33 The charge transfer state (¹CT) decayed with a time constant of 3.7 ns, a value that is consistent with the fluorescence lifetime measurements.

At first glance, transient difference spectra in the presence of Cu(I) revealed a similar picture compared to that in methanol alone; however, the bands at 480 and 630 nm characteristic of the ¹ET state appeared with substantially reduced intensity (Figure 5c). As illustrated with the transient absorption trace at 480 nm, the ¹ET state seems to evolve with similarly rapid kinetics compared to the probe in methanol alone, an observation that contradicts the strong fluorescence enhancement upon Cu(I) coordination. Given the triexponential decay kinetics of the fluorescence emission, it was not surprising that the transient absorption data did not fit to a model solely based on consecutive first-order reactions. To account for the multiexponential decay,

⁽³¹⁾ Blair, J. T.; Sahyun, M. R. V.; Sharma, D. K. J. Photochem. Photobiol.,

<u>A</u> 1994, 77, 133. Yang, G.; Wu, H. <u>J. Photochem. Photobiol., A</u> 1992, 66, 69.

⁽³³⁾ Güsten, H.; Heinrich, G.; Frühbeis, H. Ber. Bunsen Ges. Phys. Chem. 1977, 81, 810. Yan, Z. L.; Wu, S. K. J. Lumin. 1993, 54, 303. Rurack, K.; Bricks, J. L.; Schulz, B.; Maus, M.; Reck, G.; Resch-Genger, U. Phys. Chem. A 2000, 104, 6171.

Table 3. Pump-Probe Transient Absorption Data for Compounds 1a and 2 in Methanol (298 K)

cmpd	conditions	$k_1 (\tau_1)$	$k_2(\tau_2)$	$k_3(\tau_3)$	k_4 (au_4)
1a	MeOH	$1.6 \times 10^{12} (0.6 \text{ps})$	$4.2 \times 10^{10} (24 \mathrm{ps})$	$1.3 \times 10^{10} (77 \text{ ps})$	
1a	TFA (180 mM) in MeOH	$9.1 \times 10^{11} (1.1 \text{ ps})$	$1.4 \times 10^{10} (71 \text{ ps})$	$9.4 \times 10^9 (0.11 \text{ns})$	$2.7 \times 10^8 (3.7 \text{ ns})$
1a	$[Cu(I)(CH_3CN)_4]PF_6$ (10 μ M) in MeOH	$1.5 \times 10^{12} (0.7 \mathrm{ps})$	$1.2 \times 10^{10} (83 \mathrm{ps})$	$1.9 \times 10^{10} (53 \text{ ps})$	$3.1 \times 10^8 (3.2 \text{ns})$
		_	$1.3 \times 10^9 (0.77 \text{ns})$	_	
			$9.4 \times 10^9 (0.11 \text{ns})$		
2	MeOH	$1.3 \times 10^{12} (0.8 \mathrm{ps})$	$1.5 \times 10^{11} (6.6 \text{ ps})$	$1.3 \times 10^{10} (77 \text{ ps})$	$2.6 \times 10^8 (3.8 \text{ns})$

we assumed a model that involves three coordination species, A, B, and C, with distinct PET dynamics:

$$FC_{A} \xrightarrow{k_{1}} {}^{1}LE_{A} \xrightarrow{k_{2a}} {}^{1}ET_{A} \xrightarrow{k_{3a}} GS_{A}$$

$$FC_{B} \xrightarrow{k_{1}} {}^{1}LE_{B} \xrightarrow{k_{2b}} {}^{1}ET_{B} \xrightarrow{k_{3b}} GS_{B}$$

$$FC_{C} \xrightarrow{k_{1}} {}^{1}LE_{C} \xrightarrow{k_{2c}} {}^{1}CT_{C} \xrightarrow{k_{4}} GS_{C}$$

$$(4)$$

If equilibration of the three species is slow with regard to the nanosecond time scale of the experiment, each deactivation pathway can be modeled with an independent set of differential equations (see Supporting Information). Species A and B undergo PET quenching with rates k_{2a} and k_{2b} , corresponding to the two fast decay components of the fluorescence emission. Both species are assumed to form the vibrationally relaxed excited state ¹LE from the Franck-Condon state FC with the same rate constant k_1 . Furthermore, the two charge recombination rate constants k_{3a} and k_{3b} are also assumed to be identical. Species C corresponds to the slow component that involves formation of the radiative charge transfer state ¹CT. The vibrational cooling rate constant k_1 is again assumed to be identical with the other two species. The rate constant k_{2c} was obtained from the delayed fluorescence emission rate, and k_4 from the slow component of the fluorescence decay. Because neither the thiazacrown nor its Cu(I) complex absorb within the wavelength range of the acquired transients, we can assume that the extinction coefficients for the corresponding intermediates for each species remain identical. Furthermore, the triexponential decay profile implies that the three species equilibrate substantially slower than the time scale of radiative deactivation. Therefore, the concentration of the three species at time zero corresponds to the fractional amplitudes obtained from the fluorescence lifetime analysis (Table 2). Because the rate constants k_{2a} , k_{2b} , k_{2c} , and k_4 are given by the corresponding inverse lifetimes of the individual components, only k_1 , k_3 , and the four molar extinction coefficients were used as variables for nonlinear least-squares fitting. Despite the considerable constraints imposed by the fixed rate constants, the model fit very well with the data as illustrated with the single time trace at 480 nm (Figure 5c). The inverse rate constant for conversion of FC to LE was found to be 0.7 ps and compared well with the data in neutral and acidic methanol. Similarly, the charge recombination time constant of 56 ps agreed well with the previous fits. The deconvoluted transient absorption traces at 480 nm for each of the three species are shown as dotted lines in Figure 5c. In agreement with the fluorescence lifetime data, the pump-probe experiments indicate the presence of three species with distinct PET quenching kinetics. Most importantly, in the presence of Cu(I) we found no evidence for the formation of an additional transient reaction product that was not also formed in methanol alone and under acidic conditions. Consistent with the steady-state fluorescence emission profiles, the nature of all involved transient species remains qualitatively unchanged; only the kinetics of their interconversion differs as a function of Cu(I) or proton concentration.

Variable Temperature ¹H NMR Spectra. Although the fluorescence molar-ratio titrations firmly established a 1:1 stoichiometry for the interaction of Cu(I) with 1a or ligand 3, the fluorescence lifetime and pump-probe data unequivocally support the presence of three distinct species. Furthermore, given the uniform nature of the observed transient reaction products, we concluded that these three species must somehow differ with regard to their coordination mode only, for example through binding of a counterion or solvent molecule in the first coordination sphere. To find supporting evidence for this hypothesis, we conducted variable temperature ¹H NMR experiments. The initial molar-ratio titration of 3 with Cu(I) at room temperature indicated a fast exchange equilibrium with regard to the NMR time scale. If there are multiple equilibria present involving several distinct coordination species, changes in temperature should give rise to nonuniform chemical shift changes. Figure 7 shows representative ¹H NMR traces of the aromatic and aliphatic region of ligand 3 in the presence of 1 molar equiv of [Cu(I)(CH₃CN)₄]PF₆ in CD₃OD at three temperatures. Compared to the uncoordinated ligand, which is shown at the bottom, all resonances are shifted downfield. When the temperature is decreased from 20 to -20 °C, the aromatic resonances of the Ha/Ha', Hb/Hb', and Hc protons gradually drift back toward the respective positions of the free ligand (Figure 7a, left). While the chemical shift itself, even in the absence of a dynamic equilibrium, is sensitive to temperature variations, the observed changes are larger than expected. This is in fact directly confirmed by the insignificant shift of the thioether methylene resonances in the aliphatic region (Figure 7a, right). A plot of the chemical shift changes as a function of temperature best illustrates the dramatically different behavior of the two sets of proton resonances (Figure 7b). While the binding affinity of chelate complexes is expected to decrease with decreasing temperature, we would anticipate a gradual shift of all resonances upon decomplexation. The temperature insensitivity of the chemical shift for the thioether methylene protons therefore suggests that Cu(I) remains coordinated. At the same time, weakening of the nitrogen—Cu(I) interaction at lower temperatures would be expected to enhance the π -donation ability of the aniline nitrogen due to an increasingly flattened geometry with enhanced sp² character. As a consequence, the proton resonances are expected to undergo an upfield shift, as was indeed observed. Even at the lowest accessible temperature in CD₃OD, the equilibrium system did not approach the slow exchange regime or coalescence point, which indicates a fast dynamic exchange between these species. It is important to note that the time scale of NMR is vastly different compared to fluorescence spectroscopy; therefore, the fast exchange dynamics does not contradict the triexponential decay profile observed in

the lifetime measurements. The observed temperature dependence of the chemical shift changes is consistent with an equilibrium that involves at least two types of Cu(I) coordination species. Besides the expected 4-coordinate complex with the thiazamacrocycle, an additional species containing an auxiliary ligand, either a solvent molecule or possibly the counterion, must be present. As the temperature is lowered, the equilibrium is shifted in favor of the latter species due to the decreasing contribution of the entropy term $(T\Delta S)$. This in turn implies that the mixed coordination species is enthalpically more favorable compared to the tetradentate macrocyclic complex.

Quantum Chemical Calculations. Due to the rapid exchange dynamics, the variable temperature NMR experiments prevented us from identifying the individual components of the equilibrium system. To gain further insights into the nature of possible coordination species containing an auxiliary ligand, we performed a series of quantum chemical calculations. To assess the stability of the Cu(I) complex of ligand 3 coordinated to an additional solvent ligand, we used density functional theory to calculate the gas phase enthalpy $\Delta H_{\rm aux}$ of the following equilibrium (eq 5):

$$[3-Cu(I)]^{+} + solv \Rightarrow [3-Cu(I)(solv)]^{+} \Delta H_{aux}$$
 (5)

Table 4 shows all pertinent data obtained from these calculations. The lowest energy structure of complex [3-Cu(I)]⁺ assumes an all-trans t,t,t configuration, 22,34 in which all of the 6-membered chelate rings adopt a chair conformation (Figure 8, left; see also Supporting Information for more detailed structural data). Replacement of the aniline nitrogen donor with methanol yielded the ternary complex t,t-[3-Cu(I)(MeOH)]⁺, which revealed a hydrogen bonding interaction between the OH group of methanol and the aniline nitrogen (Figure 8, middle; Supporting Information). The gas phase enthalpy for formation of t,t-[3-Cu(I)(MeOH)]⁺ as defined by the equilibrium in eq 5 is exothermic with $\Delta H_{\text{MeOH}} = -12 \text{ kcal/mol}$ (Table 4), a result that is consistent with the proposed coordination of methanol at low temperature in the NMR study. If the configuration of the complex is switched from trans, trans to trans, cis the aniline nitrogen is pushed further away from the copper center and is no longer accessible for H-bonding (Figure 8, right). The geometry optimized structure of this complex c,t-[3-Cu(I)-(MeOH)]⁺ was found to be slightly higher in energy than the

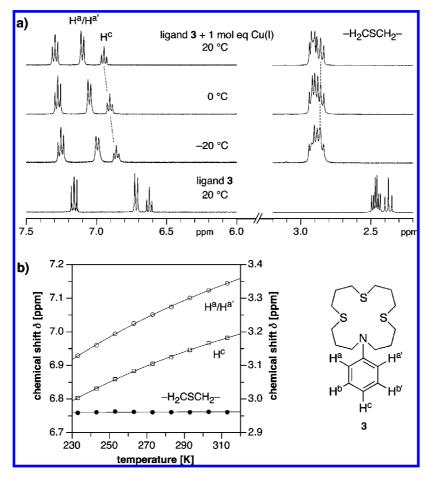


Figure 7. Variable temperature ¹H NMR experiment of ligand 3 in the presence of 1 molar equiv of [Cu(I)(CH₃CN)₄]PF₆ in CD₃OD.

Table 4. Computational Data for the Geometry Optimized Cu(I) Complexes of Ligand 3 and Methanol as an Auxiliary Ligand^a

species	Σ^b	$\Delta H_{\rm aux}{}^c$ (kcal/mol)	HOMO energy (eV)	donor obital energy ^d (eV)
t,t,t-[3-Cu(I)] ⁺	326.7	0	-8.239	-9.746
t,t-[3-Cu(I)(MeOH)] ⁺	336.7	-12.4	-8.306	-9.001
c,t-[3-Cu(I)(MeOH)] ⁺	351.0	-10.2	-7.824	-7.824

^a B3LYP/6-31+G(d,p)//B3LYP/6-31G(d) level of theory. ^b Sum of bond angles at aniline nitrogen N1 (C7-N1-C8, C8-N1-C11, C7-N1-C11; the atom numbering scheme is provided in the Supporting Information). ^c Association enthalpy as defined by eq 5. ^d homologous to aniline HOMO with B₂ symmetry.

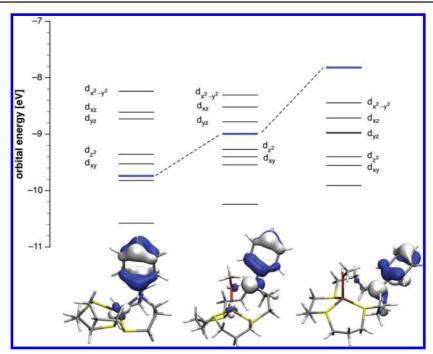


Figure 8. Effect of solvent coordination on the orbital energy levels of Cu(I) complexes with ligand 3. The depicted orbital diagrams refer to the corresponding energy levels marked in blue (see text for details).

H-bonded structure; however, the gas phase association enthalpy remained exothermic with $\Delta H_{\text{MeOH}} = -10$ kcal/mol.

Coordination of methanol to [3-Cu(I)]⁺ imposes significant structural changes that directly affect the PET thermodynamics. Judging from the sum of the bond angles at the aniline nitrogen $(\Sigma, \text{ Table 4})$, solvent coordination results in flattening of the macrocycle geometry at this position. In the parent complex [3-Cu(I)]⁺, the aniline nitrogen is sp³ hybridized and adopts a near ideal trigonal pyramidal geometry with $\Sigma = 331^{\circ}$. While in the H-bonded complex t,t-[3-Cu(I)(MeOH)]⁺ the aniline nitrogen is still somewhat pyramidalized, the increased Σ of 337° implies some flattening and therefore increased p-character of the nitrogen lone pair. In the cis, trans isomer c,t-[3-Cu(I)(MeOH)]⁺, the aniline nitrogen is engaged in neither metal coordination nor H-bonding, and assumes a near trigonal planar geometry with $\Sigma = 351^{\circ}$. With the change from a trigonal pyramidal to a trigonal planar geometry, the aniline nitrogen is transformed from a strong σ -acceptor into a π -donor. Previous quantum chemical calculations showed that the aniline molecular orbital with B2 symmetry serves as the electron donor in the PET process.¹³ As illustrated in Figure 8, the energy of this orbital is strongly affected by the structural changes imposed on the aniline nitrogen. As the aniline nitrogen approaches a more flattened geometry, its π -donor strength increases, which in turn results in an increased MO energy and therefore more favorable electron transfer thermodynamics. At the same time, the energy of the highest occupied metal-centered d-orbital remains largely unchanged, a result that strongly contrasts the widely different deactivation rates extracted from the fluorescence lifetime data of 1a in the presence of Cu(I) (Table 2). The experimental data are therefore best reconciled with a model that entails three distinct coordination species with varying ability to allow PET from the aniline B₂ MO as the electron donor. To further support this hypothesis, we repeated the above time-resolved fluorescence decay measurements in dichloromethane, a noncoordinating solvent in which no solventassociated species should form. The decay profile of 1a saturated with Cu(I) indeed fit best a monoexponential rate law with a long lifetime of $\tau = 4.03$ ns (Figure S4d, Supporting Information), thus confirming the absence of a Cu(I)-promoted quenching channel.

In summary, Figure 9 illustrates the relevant excited-state processes and the associated dynamics found for the various solution conditions. In neutral methanol, the initially formed Franck—Condon state (FC) undergoes vibrational cooling to a local excited singlet state ¹LE, which is rapidly converted to the electron transfer state ¹ET followed by charge recombination back to the initial ground state GS (Figure 9, center). Under strongly acidic conditions, the aniline nitrogen is protonated which effectively inhibits the PET process (Figure 9, left). The initially formed local excited state ¹LE is now converted to a more polarized (pyrazoline centered) charge-transfer state, which undergoes radiative deactivation back to the ground state. Finally, saturation of **1a** with Cu(I) in neutral methanol leads to the formation of three coordination species with distinct deactivation pathways (Figure 9, right).

While according to the derived model all species form a similar local excited state ¹LE, the subsequent deactivation pathways differ depending on the energy level of the ligandcentered MO acting as the electron donor in PET. In the NS₃coordinated, all-trans configured complex t,t,t-[1a-Cu(I)]⁺ (A), the energetics for PET are unfavorable and the initially formed ¹LE state decays through ¹CT with fluorescence emission back to the ground state. This complex is in equilibrium with two additional solvent coordinated species, assigned as t,t-[1a-Cu(I)(MeOH)⁺ (B) and c,t-[1a-Cu(I)(MeOH)]⁺ (C), which based on the gas phase quantum chemical calculations are expected to have a more favorable PET driving force. Although all three complexes are engaged in a dynamic exchange equilibrium that exceeds the NMR time scale, the fluorescence decay and pump-probe data can be modeled as a superimposition of the individual nonequilibrating decay pathways.

Contrary to the notion that Cu(I) might engage in reductive electron transfer quenching, we found no evidence for formation

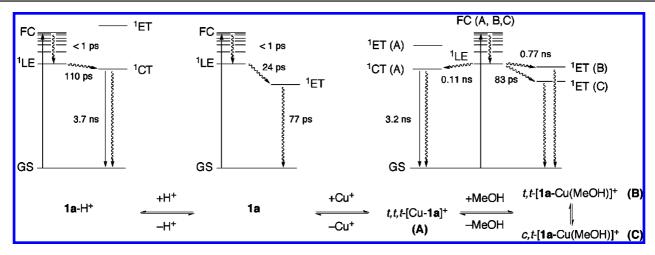


Figure 9. Jablonski diagram for the excited-state deactivation pathways of probe **1a** in neutral methanol (center), under acidic conditions (left), and in the presence of [Cu(I)(CH₃CN)₄]PF₆ (right). The indicated lifetime data are based on the pump-probe experiments (GS = ground state, FC = Franck-Condon excited state, ¹LE = local excited state, ¹ET = electron transfer state, ¹CT = emissive charge transfer excited state).

of a transient Cu(II) species upon photoexcitation of Cu(I)saturated 1a. Such a quenching pathway would require electron transfer kinetics comparable to the observed excited-state deactivation rate of $k_0 = 2.6 \times 10^8 \text{ s}^{-1}$. While many efforts have been made to generate Cu(II/I) complexes with large selfexchange electron transfer rate constants, only a few systems were described that exceeded 10⁵ M⁻¹ s⁻¹. The structurally related copper complex of 1,5,9,13-tetrathiacyclohexadecane ([16]aneS₄), which has the same ring size as ligand 3, exhibited a comparatively slow self-exchange rate with $k_{11} = 3.7 \times 10^3$ M⁻¹ s⁻¹. ^{35,36} The reorganizational energy barriers for Cu(II/I) systems are typically high due to large changes brought about the inner-coordination sphere of copper, often involving bond breakage or formation. Furthermore, the large distance between the Cu(I) center and the fluorophore also renders PET from Cu(I) less favorable. As a result, PET quenching is kinetically controlled and determined by the energetics of the aniline donor but not the Cu(I) center. Most importantly, the redox stability of the Cu(II/I) couple is not an impediment for the design of Cu(I)-selective turn-on sensors. If the solution equilibria were devoid of ternary solvent adducts and included only the binary complex t,t,t-[1a-Cu(I)]⁺ (A), the fluorescence enhancement of 1a would increase from 21- to 90-fold as judged by the ratio of the excited-state deactivation rate constants of the free ligand and the slowest component in the presence of Cu(I).

Conclusions

Following the previously reported approach for tuning of the PET thermodynamics in triarylpyrazoline-based probes, ¹² we

successfully optimized the contrast ratio of a Cu(I)-responsive fluorescent probe. While we achieved a strong 29-fold fluorescence enhancement upon saturation with Cu(I) in methanol, the contrast ratio remained significantly lower compared to acidic conditions. Detailed time-resolved spectroscopic studies revealed that the contrast ratio for Cu(I)-binding is not compromised by reductive electron transfer from Cu(I) but by the formation of several ternary coordination species with varying PET driving forces. Most importantly, the combined photophysical, electrochemical, and solution chemistry experiments suggest that electron transfer from Cu(I) does not compete with radiative deactivation of the excited fluorophore, and hence, that the Cu(I)-induced fluorescence switching is kinetically controlled. The results of this study highlight the challenges associated with designing high-contrast ratio probes for Cu(I), which not only require careful tuning of the PET thermodynamics but also careful control of the Cu(I) solution chemistry. The conclusions derived from this study are not limited to pyrazoline-based probes but are generally applicable to the design of Cu(I)selective probes composed of a rigid, electronically decoupled receptor unit attached to any other fluorophore platform.

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Supporting Information Available: Synthesis and spectroscopic characterization of derivatives 1–6 and additional information as mentioned in the text. This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽³⁴⁾ McCrindle, R.; Ferguson, G.; McAlees, A. J.; Parvez, M.; Ruhl, B. L.; Stephenson, D. K.; Wieckowski, T. *Dalton Trans.* **1986**, 2351.

⁽³⁵⁾ Rorabacher, D. B. Chem. Rev. 2004, 104, 651.

⁽³⁶⁾ Martin, M. J.; Endicott, J. F.; Ochrymowycz, L. A.; Rorabacher, D. B. *Inorg. Chem.* 1987, 26, 3012.